Package 'STITCH'

May 26, 2016

Type Package

Title STITCH - Sequencing To Imputation Through Constructing Haplotypes

Version 1.1.3

Date 2016-05-26

Author Robert William Davies

Maintainer Robert William Davies <robertwilliamdavies@gmail.com>

Description STITCH performs imputation of individuals sequenced to low coverage in a read aware fashion without a reference panel.

Installation To install, first install dependencies, then run the install.packages command, pointing to the downloaded tarball (STITCH.tar.gz)

Getting started A minimum run requires the following options to be set: the chromosome being run (chr); a path to a file with a set of bi-allelic SNP sites (posfile); a choice of K, the number of internally modelled haplotypes (K); a path to an output directory (outputdir); a path to a temporary directory, ideally on fast disks or a RAM disk (tempdir); a list of bam files (bamlist); and the number of generations since founding (nGen), which can be approximated from a choice of K for wild populations from 4 * Ne / K. Additional useful options relate to what region to impute (regionStart, regionEnd, buffer), whether to use validation data to benchmark imputation (genfile), the number of cores to use (nCores), whether imputation is run on a server or cluster (environment), the number of EM iterations (niterations), whether to run in diploid or pseudoHaploid mode (method), and if run in pseudoHaploid mode, what iteration to switch from pseudoHaploid to diploid (switchModelIteration).

Depends parallel, Rsamtools

Imports Rcpp

LinkingTo Rcpp, RcppArmadillo

RoxygenNote 5.0.1

License See attached LICENSE file

NeedsCompilation yes

5

R topics documented:

STITCH	 				•	•	•		•	•	•	•			•	•	•	•		•	•	2

Index

STITCH

Sequencing To Imputation Through Constructing Haplotypes

Usage

```
STITCH(chr, nGen = "", posfile, K, outputdir, tempdir, bamlist = "",
genfile = "", method = "diploid", outputInputInVCFFormat = FALSE,
downsampleToCov = 50, downsampleFraction = 1, readAware = TRUE,
chrStart = NA, chrEnd = NA, regionStart = NA, regionEnd = NA,
buffer = NA, maxDifferenceBetweenReads = 1000,
alphaMatThreshold = 1e-04, emissionThreshold = 1e-04,
iSizeUpperLimit = as.integer(600), bqFilter = as.integer(17),
niterations = 40, shuffleHaplotypeIterations = c(4, 8, 12, 16),
splitReadIterations = 25, nCores = 1, expRate = 0.5, maxRate = 100,
minRate = 0.1, Jmax = 100, regenerateInput = TRUE,
originalRegionName = NA, keepInterimFiles = FALSE, keepTempDir = FALSE,
environment = "server", pseudoHaploidModel = 9, diploidModel = 2,
outputHaplotypeProbabilities = FALSE, switchModelIteration = NA,
generateInputOnly = FALSE, restartIterations = NA,
refillIterations = c(6, 10, 14, 18), downsampleSamples = 1,
downsampleSamplesKeepList = NA, subsetSNPsfile = NA,
useSoftClippedBases = FALSE, outputBlockSize = 1000)
```

Arguments

chr	What chromosome to run. Should match BAM headers
posfile	Where to find file with positions to run. File is tab seperated with no header, one row per SNP, with col 1 = chromosome, col 2 = physical position (sorted from smallest to largest), col 3 = reference base, col 4 = alternate base. Bases are capitalized. Example first row: $1 < tab>1000 < tab>A < tab>G < tab>$
К	How many founder / mosaic haplotypes to use
outputdir	What output directory to use
tempdir	What directory to use as temporary directory. If possible, use ramdisk, like /dev/shm/
bamlist	Path to file with bam file locations. File is one row per entry, path to bam files. Bam index files should exist in same directory as for each bam, suffixed either .bam.bai or .bai
genfile	Path to gen file with high coverage results. Empty for no genfile. File has a header row with a name for each sample, matching what is found in the bam file. Each subject is then a tab seperated column, with $0 = \text{hom ref}$, $1 = \text{het}$, $2 = \text{hom alt and NA}$ indicating missing genotype, with rows corresponding to rows of the posfile. Note therefore this file has one more row than posfile which has no header

STITCH

method	How to run imputation - either diploid or pseudoHaploid, the former being the original method quadratic in K, the later being linear in K						
outputInputInVC	FFormat						
	Whether to output the input in vcf format						
downsampleToCov	,						
	What coverage to downsample individual sites to. This ensures no floating point errors at sites with really high coverage						
downsampleFract							
	Downsample BAMs by choosing a fraction of reads to retain. Must be value 0 <downsamplefraction<1< td=""></downsamplefraction<1<>						
readAware	Whether to run the algorithm is read aware mode. If false, then reads are split into new reads, one per SNP per read						
chrStart	When loading from BAM, some start position, before SNPs occur. Default NA will infer this from either regionStart, regionEnd and buffer, or posfile						
chrEnd	When loading from BAM, some end position, after SNPs occur. Default NA will infer this from either regionStart, regionEnd and buffer, or posfile						
regionStart	When running imputation, where to start from. The 1-based position x is kept if regionStart <= x <= regionEnd						
regionEnd	When running imputation, where to stop						
buffer	Buffer of region to perform imputation over. So imputation is run form regionStart- buffer to regionEnd+buffer, and reported for regionStart to regionEnd, including the bases of regionStart and regionEnd						
maxDifferenceBe	tweenReads						
	How much of a difference to allow the reads to make in the forward backward probability calculation. For example, if P(read state 1)=1 and P(read state 2)=1e-6, re-scale so that their ratio is this value. This helps prevent any individual read as having too much of an influence on state changes, helping prevent against influence by false positive SNPs						
alphaMatThresho							
	Minimum (maximum is 1 minus this) state switching into probabilities						
emissionThresho							
	Emission probability bounds. emissionThreshold $< P(alt read state k) < (1-emissionThreshold)$						
iSizeUpperLimit							
	Do not use reads with an insert size of more than this value						
bqFilter	Minimum BQ for a SNP in a read. Also, the algorithm uses bq<=mq, so if mapping quality is less than this, the read isnt used						
niterations	Number of EM iterations						
shuffleHaplotyp							
splitReadIterat	Iterations on which to perform heuristic attempt to shuffle founder haplotypes for better fit. To disable set to NA.						
	Iterations to try and split reads which may span recombination breakpoints for a better fit						
nCores	How many cores to use						
expRate	Expected recombination rate in cM/Mb						
maxRate	Maximum recomb rate cM/Mb						

minRate	Minimum recomb rate cM/Mb
Jmax regenerateInput	Maximum number of SNPs on a read
	Whether to regenerate input files
originalRegionN	
	If regenerateInput is FALSE (i.e. using existing data), this is the name of the original region name (chr.regionStart.regionEnd). This is necessary to load past variables
keepInterimFile	
	Whether to keep interim parameter estimates
keepTempDir	Whether to keep files in temporary directory
environment	Whether to use server or cluster multicore options
pseudoHaploidMo	del
	How to model read probabilities in pseudo diploid model (shouldn't be changed)
diploidModel outputHaplotype	How to model read probabilities in diploid model (shouldn't be changed) Probabilities
	Whether to output haplotype probabilities in files
switchModelIter	ation
	Whether to switch from pseudoHaploid to diploid and at what iteration (NA for no switching)
generateInputOn	
	Whether to just generate input data then quit
restartIteratio	
C:1174	In pseudoHaploid method, which iterations to look for collapsed haplotype prnob- abilities to resolve
refillIteration	
downsampleSampl	When to try and refill some of the less frequently used haplotypes
downsampresampr	What fraction of samples to retain. Useful for checking effect of N on imputa-
doumoomploCompl	tion. Not meant for general use
downsampleSampl	When downsampling samples, specify a numeric list of samples to keep
	If input data has already been made for a region, then subset down to a new set of SNPs, as given by this file. Not meant for general use
useSoftClippedB	
outputBlockSize	Whether to use (TRUE) or not use (FALSE) bases in soft clipped portions of reads
Outputbiock312e	
	How many samples to write out to disk at the same time when making temporary VCFs that are later pasted together at the end to make the final VCF. Smaller means lower RAM footprint, larger means faster write.
Number	of generations since founding or mixing. Note that the algorithm is relatively robust to this. Use $nGen = 4 * Ne / K$ if unsure

Value

Results in properly formatted version

Author(s)

Robert Davies

Index

STITCH, 2