Package 'STITCH'

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Type Package

Title STITCH - Sequencing To Imputation Through Constructing Haplotypes

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Description STITCH performs imputation of individuals sequenced to low coverage in a read aware fashion without a reference panel.

Installation To install, first install dependencies, then run the install.packages command, pointing to the downloaded tarball (STITCH.tar.gz)

Getting started A minimum run requires the following options to be set: the chromosome being run (chr); a path to a file with a set of bi-allelic SNP sites (posfile); a choice of K, the number of internally modelled haplotypes (K); a path to an output directory (outputdir); a path to a temporary directory, ideally on fast disks or a RAM disk (tempdir); a list of bam files (bamlist); and the number of generations since founding (nGen), which can be approximated from a choice of K for wild populations from 4 * Ne / K. Additional useful options relate to what region to impute (regionStart, regionEnd, buffer), whether to use validation data to benchmark imputation (genfile), the number of cores to use (nCores), whether imputation is run on a server or cluster (environment), the number of EM iterations (niterations), whether to run in diploid or pseudoHaploid mode (method), and if run in pseudoHaploid mode, what iteration to switch from pseudoHaploid to diploid (switchModelIteration).

Depends parallel, Rsamtools

Imports Rcpp

LinkingTo Rcpp, RcppArmadillo

RoxygenNote 5.0.1

License See attached LICENSE file

NeedsCompilation yes

R topics documented:

STITCH   *Sequencing To Imputation Through Constructing Haplotypes*

Usage

```
STITCH(chr, nGen = "", posfile, K, outputdir, tempdir, bamlist = "",
  cramlist = "", reference = "", genfile = "", method = "diploid",
  outputInputInVCFFormat = FALSE, downsampleToCov = 50,
  downsampleFraction = 1, readAware = TRUE, chrStart = NA, chrEnd = NA,
  regionStart = NA, regionEnd = NA, buffer = NA,
  maxDifferenceBetweenReads = 1000, alphaMatThreshold = 1e-04,
  emissionThreshold = 1e-04, iSizeUpperLimit = as.integer(600),
  bqFilter = as.integer(17), niterations = 40,
  shuffleHaplotypeIterations = c(4, 8, 12, 16), splitReadIterations = 25,
  nCores = 1, expRate = 0.5, maxRate = 100, minRate = 0.1, Jmax = 100,
  regenerateInput = TRUE, originalRegionName = NA,
  keepInterimFiles = FALSE, keepTempDir = FALSE, environment = "server",
  pseudoHaploidModel = 9, diploidModel = 2,
  outputHaplotypeProbabilities = FALSE, switchModelIteration = NA,
  generateInputOnly = FALSE, restartIterations = NA,
  refillIterations = c(6, 10, 14, 18), downsampleSamples = 1,
  downsampleSamplesKeepList = NA, subsetSNPsfile = NA,
  useSoftClippedBases = FALSE, outputBlockSize = 1000)
```

Arguments

chr	What chromosome to run. Should match BAM headers
posfile	Where to find file with positions to run. File is tab seperated with no header, one row per SNP, with col 1 = chromosome, col 2 = physical position (sorted from smallest to largest), col 3 = reference base, col 4 = alternate base. Bases are capitalized. Example first row: 1<tab>1000<tab>A<tab>G<tab>
K	How many founder / mosaic haplotypes to use
outputdir	What output directory to use
tempdir	What directory to use as temporary directory. If possible, use ramdisk, like /dev/shm/
bamlist	Path to file with bam file locations. File is one row per entry, path to bam files. Bam index files should exist in same directory as for each bam, suffixed either .bam.bai or .bai
cramlist	Path to file with cram file locations. File is one row per entry, path to cram files. cram files are converted to bam files on the fly for parsing into STITCH
reference	Path to reference fasta used for making cram files. Only required if cramlist is defined

genfile   Path to gen file with high coverage results. Empty for no genfile. File has a header row with a name for each sample, matching what is found in the bam file. Each subject is then a tab seperated column, with 0 = hom ref, 1 = het, 2 = hom alt and NA indicating missing genotype, with rows corresponding to rows of the posfile. Note therefore this file has one more row than posfile which has no header

method   How to run imputation - either diploid or pseudoHaploid, the former being the original method quadratic in K, the later being linear in K

outputInputInVCFFormat
                    Whether to output the input in vcf format

downsampleToCov
                    What coverage to downsample individual sites to. This ensures no floating point errors at sites with really high coverage

downsampleFraction
                    Downsample BAMs by choosing a fraction of reads to retain. Must be value 0<downsampleFraction<1

readAware   Whether to run the algorithm is read aware mode. If false, then reads are split into new reads, one per SNP per read

chrStart   When loading from BAM, some start position, before SNPs occur. Default NA will infer this from either regionStart, regionEnd and buffer, or posfile

chrEnd   When loading from BAM, some end position, after SNPs occur. Default NA will infer this from either regionStart, regionEnd and buffer, or posfile

regionStart   When running imputation, where to start from. The 1-based position x is kept if regionStart <= x <= regionEnd

regionEnd   When running imputation, where to stop

buffer   Buffer of region to perform imputation over. So imputation is run form regionStart-buffer to regionEnd+buffer, and reported for regionStart to regionEnd, including the bases of regionStart and regionEnd

maxDifferenceBetweenReads
                    How much of a difference to allow the reads to make in the forward backward probability calculation. For example, if P(read | state 1)=1 and P(read | state 2)=1e-6, re-scale so that their ratio is this value. This helps prevent any individual read as having too much of an influence on state changes, helping prevent against influence by false positive SNPs

alphaMatThreshold
                    Minimum (maximum is 1 minus this) state switching into probabilities

emissionThreshold
                    Emission probability bounds. emissionThreshold < P(alt read | state k) < (1-emissionThreshold)

iSizeUpperLimit
                    Do not use reads with an insert size of more than this value

bqFilter   Minimum BQ for a SNP in a read. Also, the algorithm uses bq<=mq, so if mapping quality is less than this, the read isnt used

niterations   Number of EM iterations

shuffleHaplotypeIterations
                    Iterations on which to perform heuristic attempt to shuffle founder haplotypes for better fit. To disable set to NA.

splitReadIterations

    Iterations to try and split reads which may span recombination breakpoints for a better fit

nCores	How many cores to use
expRate	Expected recombination rate in cM/Mb
maxRate	Maximum recomb rate cM/Mb
minRate	Minimum recomb rate cM/Mb
Jmax	Maximum number of SNPs on a read

regenerateInput

    Whether to regenerate input files

originalRegionName

    If regenerateInput is FALSE (i.e. using existing data), this is the name of the original region name (chr.regionStart.regionEnd). This is necessary to load past variables

keepInterimFiles

    Whether to keep interim parameter estimates

keepTempDir	Whether to keep files in temporary directory
environment	Whether to use server or cluster multicore options

pseudoHaploidModel

    How to model read probabilities in pseudo diploid model (shouldn't be changed)

diploidModel	How to model read probabilities in diploid model (shouldn't be changed)

outputHaplotypeProbabilities

    Whether to output haplotype probabilities in files

switchModelIteration

    Whether to switch from pseudoHaploid to diploid and at what iteration (NA for no switching)

generateInputOnly

    Whether to just generate input data then quit

restartIterations

    In pseudoHaploid method, which iterations to look for collapsed haplotype prnobabilities to resolve

refillIterations

    When to try and refill some of the less frequently used haplotypes

downsampleSamples

    What fraction of samples to retain. Useful for checking effect of N on imputation. Not meant for general use

downsampleSamplesKeepList

    When downsampling samples, specify a numeric list of samples to keep

subsetSNPsfile	If input data has already been made for a region, then subset down to a new set of SNPs, as given by this file. Not meant for general use

useSoftClippedBases

    Whether to use (TRUE) or not use (FALSE) bases in soft clipped portions of reads

outputBlockSize

    How many samples to write out to disk at the same time when making temporary VCFs that are later pasted together at the end to make the final VCF. Smaller means lower RAM footprint, larger means faster write.

Number	of generations since founding or mixing. Note that the algorithm is relatively robust to this. Use nGen = 4 * Ne / K if unsure

Value

Results in properly formatted version

Author(s)

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